# KEVIN YAMAUCHI

#### **EDUCATION**

# Ph.D. Bioengineering – UC Berkeley

2017

NSF Graduate Research Fellow

Dissertation: "Microfluidic tools for high throughput, multiparameter analysis of single cells"

M.S. Mechanical Engineering – California Polytechnic State University, San Luis Obispo 2012 NSF Graduate Research Fellow

Thesis: "Prediction of Articular Cartilage Remodeling During Dynamic Compression with a Finite Element Model"

B.S. Mechanical Engineering – California Polytechnic State University, San Luis Obispo 2012 Focus: Mechatronics/controls

Study Abroad – Hochschule München, Munich, Germany

Sept. 2009 - March 2010

Email: kevin.yamauchi@gmail.com

#### EXPERIENCE

# Senior R&D Engineer, Chan Zuckerberg Biohub

July 2017 - Present

- Collaborated with biologists to build tools that enable them to ask new questions.
- Designed and validated an automated in situ transcriptomics platform that is currently in use by four research groups.
- Wrote image processing pipelines for in situ transcriptomics including spot detection/decoding and cell segmentation (CellProfiler and Ilastik)
- Contributed to open source scientific software including napari (python n-dimensional image viewer) and starfish (python image processing toolbox for in situ transcriptomics).

#### Graduate Student Researcher, Advisor: Dr. Amy Herr

May 2013 - May 2017

- Designed microfluidic tools for high-specificity single-cell protein analysis
- Invented a device for measurement of protein localization and expression in single cells without fixation
- Designed, demonstrated, and validated first reported single-cell resolution isoelectric focusing (IEF) assay capable of resolving isoforms of endogenous proteins from single cells
- Designed and fabricated pump-less PDMS microfluidic devices for chemical gradient formation
- ullet Simulated digital chromatic aberration correction with lightfield imaging (MATLAB + Zemax)
- Developed image processing software for quantifying single-cell western blotting data
- Course developer, teaching assistant, and participant in the Cold Spring Harbor Laboratories Single Cell Analysis course (2014-2016)
- Coursework in microfluidics, optical engineering, and computational biology

### Microfluidics Intern, 10X Genomics

June 2016 - Aug. 2016

- Designed, built, and tested an automated imaging system for microfluidic device characterization
- Collaborated with a cross-functional team to determine system specifications and validate system performance

# Microscopy Museum Exhibit, Advisor: Dr. Daniel Fletcher

Sept. 2013 - Present

- Designed a microscopy-based museum exhibit with automated sample recognition and assisted focus for the Deutsches Museum (Munich, Germany)
- Led the development of the sample recognition and focusing systems
- Gained experience prototyping and integrating electromechanical and optical systems

# M.S. Thesis, Advisor: Dr. Stephen Klisch

April 2011 - June 2012

- Created a finite element model of articular cartilage with custom subroutines to predict changes in glycosaminoglycan density and collagen volume fraction distribution due to mechanical stimuli
- ullet Developed a method to estimate collagen fiber distribution with quantitative polarized light microscopy

# Low-cost Prosthetic Foot Design, Advisor: Dr. Brian Self

Sept. 2010 - June 2011

- Worked with an interdisciplinary team to design a low-cost prosthetic foot for use in developing countries
- Tested the prosthetic foot onsite at Vida Nueva, a prosthesis clinic in Choluteca, Honduras

Undergraduate Researcher, Advisors: Dr. Stephen Klisch and Dr. Robert Sah Une 2009 - Sept. 2010

- Developed and carried out a study to elucidate the effects of dynamic unconfined compression on the biomechanical properties of articular cartilage
- Designed a method to study the deformation of articular cartilage during dynamic unconfined compression using optical methods

#### Mechanical Engineering Intern, Consolidated Metco

June 2008 - Aug. 2008

- Determined the cause of failure of a frame bracket (heavy duty truck) through finite element analysis and mechanical testing
- Collaborated with failure analysts and design engineers to prototype solutions to improve the fatigue resistance of the component

#### Manuscripts in Preparation

Yamauchi KA et al. InSituToolkit: modular hardware for automating in situ transcriptomics assays.

Vo-Phamhi J, **Yamauchi KA**, Gomez-Sjoberg R. Crowd-sourced annotations for optimizing in situ transcriptomics image processing parameter selection.

Linnarsson, L, Yamauchi KA, Gomez-Sjoberg R. A pipetting robot compatible flow cell for surface-based sample preparation.

Vlassakis J, **Yamauchi KA**, Herr AE. Summit: a semi-automated algorithm for high-throughput single-cell electrophoresis image analysis

Jeeawoody S, **Yamauchi KA**, Su A, Herr AE. Investigating the impact of gel microarchitecture on immunoprobed isoelectric focusing performance in thin-film gels

### **PUBLICATIONS**

Pan Q, Yamauchi KA, and Herr AE. Controlling Dispersion during Single-Cell Polyacrylamide-Gel Electrophoresis in Open Microfluidic Devices. *Analytical Chemistry*, 2018.

Yamauchi KA\*, Tentori AM\*, and Herr AE. Arrayed isoelectric focusing using photopatterned multi?domain hydrogels. *Electrophoresis*, 2018. \*equal contribution.

Abdel-Sayed P, **Yamauchi KA**, Gerver RE, Herr AE. Fabrication of an open microfluidic device for immunoblotting. *Analytical Chemistry*, 2017.

Sinkala E et al. Profiling protein expression in circulating tumor cells using microfluidic western blotting. *Nature Communications*, 2017.

Yamauchi KA and Herr AE. Subcellular Western Blotting of Single Cells. *Microsystems and Nanoengineering*, 2017.

Tentori AM\*, **Yamauchi KA\*** and Herr AE. Detection of Isoforms Differing by a Single Charge Unit in Individual Cells. *Angewandte Chemie*, 2016. \*equal contribution.

Kang CC, Yamauchi KA\*, Vlassakis J\*, Sinkala E, Duncombe T, Herr AE. Single-cell resolution Western blotting. *Nature Protocols*, 2016. \*equal contribution

Stender ME, Raub CB, **Yamauchi KA**, Shirazi R, Vena P, Sah RL, Hazelwood SJ, and Klisch SM. Integrating qPLM and biomechanical test data with an anisotropic fiber distribution model and predictions of TGF-1 and IGF-1 regulation of articular cartilage fiber modulus. *Biomechanics and Modeling in Mechanobiology*, 2012

## Presentations

Sofroniew N, Evans K, Axelrod S, Freeman J, Can Solak A, Chhun B, **Yamauchi K**, Royer L, Nunez-Iglesias J. napari: multi-dimensional image visualization in python. *BioImage Informatics 2019* (Oral)

Ganguli D, Carr A, Long B, **Yamauchi K**, Tung T, Axelrod S, Evans K, Kiggins J, Freeman J. A comparative and unifying approach to image-based transcriptomics data analysis. *BioImage Informatics 2019* (Oral)

Yamauchi KA, Tentori AM, and Herr AE. Isoform Cytometry: Isoelectric focusing resolves protein isoforms differing by a single-charge unit with single-cell resolution *The 20th International Conference on Miniaturized Systems for Chemistry and Life Sciences (microTAS2016) (Oral)* 

Yamauchi KA, Tentori AM, and Herr AE. Isoelectric focusing resolves protein isoforms differing by a single-charge unit with single-cell resolution. *International Symposium on High Performance Liquid Phase Separations and Related Techniques (HPLC2016) (Oral)* 

Yamauchi KA and Herr AE. Sub-cellular Western Blotting of Single Cells. 2016 Berkeley Stem Cell Center Retreat (Oral)

Yamauchi KA and Herr AE. Sub-cellular Western Blotting of Single Cells. The 18th International Conference on Miniaturized Systems for Chemistry and Life Sciences (microTAS2014) (Oral)

Yamauchi KA, Raub CB, Chen AC, Sah RL, Hazelwood SJ, and Klisch SM. Glycosaminoglycan and collagen remodeling during *in vitro* dynamic compression of articular cartilage: experiments and finite element modeling. 2013 ASME Summer Bioengineering Conference (Poster)

Yamauchi KA, Williams GM, Chen AC, Hazelwood SJ, Klisch SM, and Sah RL. Radial expansion of immature cartilage during cyclic unconfined compression quantified as dynamic poisson ratio. 2011 ORS Annual Meeting (Poster)

#### PATENTS AND PATENT APPLICATIONS

A.E. Herr, et al., "Electrophoresis devices and methods for making and using the same." US Patent No. US20150316547

A.E. Herr, et al., "Isoelectric Focusing Arrays and Methods of Use Thereof." PCT Application No. PCT/US2015/33375

A.E. Herr, et al., "Subcellular Western Blotting of Single Cells." US Application No. 62/217,530

#### Course Development & Teaching Experience

# Course Developer & Teaching Assistant, UCSF Fab@Lab course

2019

- Developed a three week intensive lab course teaching biology PhD students how to design, build, and test an automated luminometer
- Lectured on programming Arduino microcontrollers.
- Course evalutation: 4.8/5.0 (university and department averages are 4.3)

Course Developer & Teaching Assistant, Cold Spring Harbor Single Cell Analysis course 2015 - 2016

- Developed a hands-on course module to teach the theory and practice of the single-cell western blotting assay
- Taught the first course on single-cell western blotting to a class of 17 scientists from around the world
- Consulted with students on ways to integrate single-cell western blotting into their scientific inquiry

## Graduate Student Instructor, UC Berkeley BioE Senior Capstone Design

2014

- Taught the engineering design method to a class of 30 senior bioengineering undergraduate students
- Developed and taught a workshop on rapid prototyping of mechatronics and automation with Arduino
- Advised students on applying the engineering design process to real clinical problems

#### SKILLS

Microfluidics:

o MATLAB

#### • Assay design and validation • Mass transport Microscopy Image Processing • Microfabrication o Simulation driven design **Mechatronics:** $\circ$ Arduino/Atmel • PCB design (EAGLE) o Embedded system design Simulation and Design: • Abaqus o Pro/Engineer o Zemax OpticStudio $\circ$ COMSOL $\circ$ SolidWorks $\circ$ Rapid prototyping ( $\mu m$ - m length scale) **Programming:**

• Python

∘ C++

| Siebel Scholar  | 2016 - 2017 |
|---|-------------|
| National Science Foundation Graduate Research Fellowship  | 2011 - 2016 |
| Lloyd Graduate Fellow   | 2016        |
| Outstanding Graduate Student Instructor<br>Awarded to top 9% of teaching assistants at UC Berkeley by faculty nomination              | 2014        |
| CBMS Student/Young Researcher Grant   | 2014        |
| Outstanding Graduate Student<br>Awarded at California Polytechnic State University - San Luis Obispo to one graduate student per year | 2012        |
| Leadership and Mentorship   |             |
|   |             |

### Intern mentor, Chan Zuckerberg Biohub

2018 - present

- Mentor to an intern writing image processing software in support of the in situ transcriptomics platform. We currently have a manuscript in progress reporting our work.
- Mentor to an intern designing custom hardware for a pipetting robot. We are currently co-authoring a manuscript sharing the designs and device characterization.

#### Research mentor, UC Berkeley

2013 - Present

- Mentor to junior researchers in the Herr Lab with a focus on experimental design and quantitative measurement. Mentorship approach includes weekly one-on-one meetings, training on laboratory and data analysis methods, and iterative feedback on scientific communication.
- Mentees include:
  - Two undergraduate students through BEGROw (initiative to recruit and retain underrepresented minorities)
  - Two undergraduates students through the UC Berkeley Undergraduate Research Apprenticeship Program
  - One UC Berkely Stem Cell Center Research Fellow
  - Two graduate rotation students

Fellowship Mentor, Latino/a Association of Graduate Students in Engineering and Science 2015 - 2016

Admissions Committee, UC Berkeley/UCSF Graduate Program in Bioengineering 2014 - 2016

Leadership Team Member, Point-of-Care Diagnostics IdeaLab 2013 - Present

- Organized a one-day conference on the role of technology in global health (January 2014)
  - Curated a biweekly seminar on point-of-care diagnostics featuring invited speakers

Mentor, Big Bear Bioengineering Undergraduate Mentoring Program Volunteer, Centro de Miembros Artificiales - La Paz, Bolivia

2012 - 2013

2012

- Volunteered for two months at Centro de Miembros Artificiales, a prosthetics clinic in La Paz, Bolivia
- Assessed prosthetic foot fabrication methods and designed a new, optimized protocol
- Redesigned inventory system, reducing procurement costs and downtime due to low supplies
- Gained experience in evaluating local resources for project planning

# Engineering World Health - Cal Poly Chapter

2010 - 2012

- Senior Project Mentor: 2011-2012
- Vice President: 2010-2011Design Project Leader: 2010

Tutor, Statics, Dynamics, and Mechanics of Materials

2011 - 2012

Tutor, Mechanism Design

2009 - 2010